

Subscriber access provided by ISTANBUL TEKNIK UNIV

Labdane Diterpenes from Calceolaria densifolia

Juan A. Garbarino, and Aurora Molinari

J. Nat. Prod., 1992, 55 (6), 744-747• DOI: 10.1021/np50084a006 • Publication Date (Web): 01 July 2004

Downloaded from http://pubs.acs.org on April 4, 2009

More About This Article

The permalink http://dx.doi.org/10.1021/np50084a006 provides access to:

- Links to articles and content related to this article
- Copyright permission to reproduce figures and/or text from this article



Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

LABDANE DITERPENES FROM CALCEOLARIA DENSIFOLIA¹

JUAN A. GARBARINO*

Departamento de Química, Facultad de Ciencia, Universidad Técnica Federico, Santa María, Casilla 110-V, Valparaíso, Chile

and AURORA MOLINARI

Instituto de Química, Facultad de Ciencias Básicas y Matemática, Universidad Católica de Valparaíso, Casilla 4059, Valparaíso, Chile

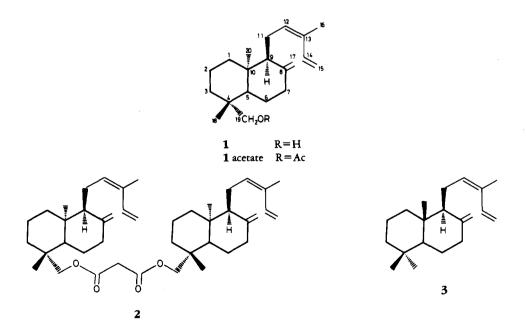
ABSTRACT.—The aerial parts of *Calceolaria densifolia* afforded two new labdane-type diterpenes, *ent-9-epi*-labda-8(17),(12Z), 14-trien-19-ol [1] and a bisditerpene 2 derived from the esterification of malonic acid by two units of the above diterpene. The structures of the new compounds have been established by spectroscopic means.

We have been studying the diterpenes present in the *Calceolaria* genus (1,2). We now report the isolation and structure elucidation of two new labdatriene diterpenes from *Calceolaria densifolia* Phil. (Scrophulariaceae), a species that grows on the hills of northern Chile (3). This is the first report of labdane-type diterpenes isolated from *Calceolaria* species.

RESULTS AND DISCUSSION

The petroleum ether extract of the aerial parts of C. densifolia, after successive cc purifications, afforded the compounds 1 and 2.

Compound 1 was purified and characterized after acetylation as 1 acetate, $C_{22}H_{34}O_2$ ([M]⁺ at m/z 330). It was an oil whose ir spectrum showed absorption bands characteristic of acetate and double bonds. Its ¹H- and ¹³C-nmr spectra displayed characteristic signals of a diterpene having a labdatriene skeleton (4–7).



¹Part 9 in the series "Diterpenoids from Calceolaria Species." For Part 8 see Chamy et al. (2).

Carbon	DEPT	Compound		
		1 acetate	2	3
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	CH_{2} Me CH_{2} Me CH_{2} Me CH_{2} Me	36.50 18.65 36.50 36.76 ^a 46.63 24.53 31.71 147.96 58.22 37.82 ^a 23.41 133.84 131.64 130.35 113.16 19.70 109.96 27.39 66.88 23.05	2 36.47 ^a 18.64 36.93 ^b 46.68 24.56 31.71 147.89 58.24 37.85 ^b 23.42 133.87 131.93 130.35 113.23 19.78 110.08 27.33 68.09 23.05	39.3 19.5 42.2 33.5 55.5 24.3 38.2 148.2 57.4 39.6 22.1 133.9 131.4 131.6 113.0 19.7 107.7 33.6 21.8 14.5
OAc	C Me	171.22 20.93		
COCH ₂ CO	C C CH ₂		166.67 166.67 41.70	

 TABLE 1.
 ¹³C-nmr Spectral Data of Compounds 1 acetate, 2, and 3 (in CDCl₃ with TMS as internal standard).

^{a,b}These values may be interchanged within the same column.

The ¹H-nmr spectrum showed signals of two quaternary methyls at δ 0.90 and 0.96. An AB system was present corresponding to an acetoxymethylene group. From the chemical shifts of -CH₂OAc (¹H nmr, two doublets at δ 3.85 and 4.23; ¹³C nmr, 66.88), a C-19 axial orientation was deduced for this grouping (8,9).

The ¹³C-nmr spectrum showed signals at δ 147.96 and 109.96 attributed, respectively, to C-8 and C-17 of the exocyclic methylene group; these were consistent with ¹H-nmr absorptions at δ 4.48 (1H, d, H-17) and 4.65 (1H, d, H-17').

The ¹H-nmr spectrum also exhibited signals at δ 5.25 (1H, t, H-12), 6.72 (1H, dd, H-14), 5.15 (1H, dd, H-15 trans), 5.05 (1H, dd, H-15 cis), and 1.75 (3H, br s, H-16), showing the presence of a Z-type conjugated diene system (4,5,10). The ¹³C-nmr signals at δ 133.84, 131.64, 130.35, and 113.16, which correspond to C-12, C-13, C-14, and C-15, respectively, were in agreement with the proposed structure for the side chain (6,7). Table 1 shows a comparison of ¹³C-nmr data with those of a known related labdane **3** (6).

The chemical shifts assigned to C-1 (δ 36.5) and to C-5 (δ 46.63) occur upfield; on the other hand, the signal attributed to C-20 (δ 23.05) appeared downfield compared. with the values observed in similar compounds (6). These effects, which are also observed in other diterpenoids obtained from the *Calceolaria* genus (11–13), are explained by the interaction imposed by a different configuration of C-11 (14) observed in the 9epimeric diterpenoids, which cause a shielding effect on C-1 and C-5 and a deshielding effect on C-20. This can be clearly demonstrated with molecular models. Therefore, the spectral data of compound **1** acetate correspond to *ent-9-epi*-labda-8(17),(12Z), 14-trien-19-yl acetate, and thereby the natural substance **1** is *ent-9-epi*-labda-8(17),(12Z), 14-trien-19-ol.

The structure of compound 2 was deduced by comparing its spectral data with those of the compound 1 acetate. In fact, the ¹H-nmr spectrum of 2 was very similar to that of 1 acetate and exhibited characteristics of the same 8(17),(12Z),14-labdatriene nucleus but with a malonate unit (δ 3.38, 2H, s) instead of an acetoxy group at C-19. However, integration due to the terpene moiety was twice that of the malonate residue, which suggested that two diterpene units must be linked by malonic acid. The ¹³C-nmr spectrum exhibited 22 signals that agreed well with those of 1 acetate. Therefore, on the basis of these data, 2 is shown to be bis-[ent-9-epi-labda-8(17),(12Z),14-trien-19-yl] malonate.

Assignments of structures and configurations for both compounds were made by comparison with spectral data of similar diterpenes (4-7) and also from biogenetic considerations (11-13).

This is the first report of the presence of labdane metabolites in the genus *Calceolaria*. These 9-epimeric diterpenes correspond to a modification of the normal diterpenoid cyclization of geranylgeranyl-pyrophosphate, which occurs in other members of the genus. The formation of these compounds might involve a chair-boat cyclization to form a bicyclic intermediate possessing an H-9 α and a 10 α -Me group (11). The presence of 9-epimeric diterpenoids and malonyl esters in species of *Calceolaria* may be of chemosystematic value.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.—¹H-nmr spectra were recorded at 250 MHz in $CDCl_3$ with TMS as internal standard; assignments of ¹³C nmr chemical shifts were made with the aid of DEPT. Mass spectra were obtained by direct inlet, 70 eV. $CHCl_3$ solutions were used for ir, and MeOH solutions for uv.

PLANT MATERIAL.—C. densifolia was collected in Parque Nacional Fray Jorge, IV Región, Chile, in September 1987. A voucher specimen (# 0138) is deposited at Universidad Técnica Federico Santa María.

EXTRACTION AND ISOLATION.—The aerial parts of *C. densifolia* (1.5 kg) were extracted at room temperature with petroleum ether for 72 h, affording 33 g of a syrup. This crude material was chromatographed on a Si gel column and eluted with petroleum ether/EtOAc mixtures of increasing polarity. Fractions of 125 ml were taken and combined based upon tlc and ¹H-nmr (60 MHz) monitoring. The intermediate fractions 13–16 provided a mixture that contained compound **2**. Fractions 26–28 gave a mixture containing compound **1**. This mixture was treated with Ac₂O/pyridine for 3 h at room temperature; the acetylated material was subjected to Si gel cc using petroleum ether-EtOAc (40:1), and fractions of 25 ml were taken. Fractions 14–17 were repurified by Si gel cc impregnated with AgNO₃ (10%), using petroleum ether-EtOAc (8:1), affording **1** acetate (210 mg).

The mixture containing **2** was subjected to Si gel using petroleum ether-EtOAc (50:1), and fractions of 25 ml were taken. Fractions 20–30, which contained **2**, were repurified by Si gel cc impregnated with AgNO₃ (10%), using petroleum ether-EtOAc (4:1), affording **2** (60 mg).

ent-9-epi-*Labda-8*(17),(12Z),14-trien-19-yl acetate [1 acetate].—Viscous yellow oil: $[\alpha]^{25}D + 23.8^{\circ}$ (CHCl₃, c = 1.85); uv λ max nm (log ϵ) 227 (4.03); ir ν max (CHCl₃) cm⁻¹ 3080, 3020, 2980–2950, 1735, 1650, 1470, 1450, 1400, 1360, 1250, 1210, 1030, 990, 900; ¹H nmr (250 MHz) δ 6.72 (1H, dd, J = 17.5, 11.0 Hz, H-14), 5.25 (1H, t, J = 7.0 Hz, H-12), 5.15 (1H, dd, J = 17.5, 1.0 Hz, H-15 trans), 5.05 (1H, dd, J = 11.0, 1.0 Hz, H-15 cis), 4.65 (1H, d, J = 2.0 Hz, H-17), 4.48 (1H, d, J = 2.0 Hz, H-17'), 4.23 (1H, d, J = 12.5 Hz, H-19), 3.85 (1H, d, J = 12.5 Hz, H-19'), 2.00 (3H, s, OAc), 1.75 (3H, br s, H-16), 0.96, 0.90 (3H, each, s, H-18, H-20); ¹³C nmr see Table 1; ms m/z (rel. int.) [C₂₂H₃₄O₂]⁺ 330 (8), [M - Me]⁺ 315 (3), [M - HOAc]⁺ 270 (7), [M - CH₂OAc]⁺ 257 (98), [M - HOAc - C₆H₉]⁺ 189 (56), [C₆H₉]⁺ 81 (100).

Bis-[ent-9-epi-labda-8(17),(12Z),14-trien-19-yl] malonate [2].—Viscous yellow oil: $[\alpha]^{25}D + 68.3^{\circ}$ (CHCl₃, c = 1.20); uv λ max nm (log ϵ) 226 (4.01); ir ν max (CHCl₃) cm⁻¹ 3080, 3020, 2980–2850, 1730, 1650, 1470, 1450, 1380, 1160, 1010, 900; ¹H nmr (250 MHz) δ 6.72 (2H, dd, J = 17.5, 11.0 Hz, H-14 and H-14a), 5.25 (2H, t, J = 7.0 Hz, H-12 and H-12a), 5.15 (2H, dd, J = 17.5, 1.0 Hz, H-15 trans and H-15a trans), 5.05 (2H, dd, J = 11.0, 1.0 Hz, H-15 cis and H-15a cis), 4.66 (2H, d, J = 2.0 Hz, H-17 and H-17a), 4.50 (2H, d, J = 2.0 Hz, H-17' and H-17'a), 4.32 (2H, d, J = 12.5 Hz, H-19 and H-19a), 3.89 (2H, d, J = 12.5 Hz, H-19' and H-19'a), 3.38 (2H, s, malonyl), 1.75 (6H, br s, H-16 and H-16a), 0.98, 0.93 (6H each, s, H-18, H-18a, H-20, and H-20a); ¹³C nmr see Table 1; ms m/z (rel. int.) $[M - C_6H_9]^+$ 563 (2), $[M - C_{20}H_{32}O]^+$ 356 (2), $[C_{20}H_{32}O]^+$ 288 (3), $[M - C_{23}H_{34}O_4]^+$ 270 (5), $[C_{19}H_{29}]^+$ 257 (14), 189 (13), 175 (20), 160 (28), 145 (100), 135 (26), 119 (32), 109 (42), 95 (48), $[C_6H_9]^+$ 81 (57), 79 (35), 67 (32).

ACKNOWLEDGMENTS

We are grateful to Professor P. Manitto (Universitá degli Studi di Milano, Italia) for recording ms, ¹H-nmr, and ¹³C-nmr spectra and to Professor O. Zoellner (Universidad Católica de Valparaíso, Chile) for the identification of plant material. This research was supported by grant No. 891303 from DGICYT, Universidad Técnica Federico Santa María, and FONDECYT No. 408/90.

LITERATURE CITED

- 1. J.A. Garbarino and A. Molinari, Phytochemistry, 29, 3040 (1990).
- M.C. Chamy, M. Piovano, J.A. Garbarino, C. Miranda, V. Gambaro, M.L. Rodriguez, C. Ruiz-Pérez, and I. Brito, *Phytochemistry*, **30**, 589 (1991).
- 3. C. Marticorena and M. Quezada, Gayana, Botánica, 42, 1 (1988).
- 4. B.R. Thomas, Acta Chem. Scand., 20, 1074 (1966).
- 5. F. Bohlmann, J. Jakupovic, A. Schuster, R. King, and H. Robinson, *Phytochemistry*, 21, 2317 (1982).
- 6. N. Noma, F. Suzuki, K. Gamou, and N. Kawashima, Phytochemistry, 21, 395 (1982).
- 7. M. Sholichin, K. Yamasaki, R. Miyama, S. Yahara, and O. Tanaka, *Phytochemistry*, 19, 326 (1980).
- 8. J. Dentali, J. Hoffmann, S. Jolad, and B. Timmermann, Phytochemistry, 26, 3025 (1987).
- 9. R.C. Cambie, J. Org. Chem., 40, 3789 (1975).
- 10. Y. Asakawa, M. Toyota, and T. Tsunematsu, Phytochemistry, 18, 1681 (1979).
- 11. M. Piovano, V. Gambaro, M.C. Chamy, J.A. Garbarino, M. Nicoletti, J. Guilhem, and C. Pascard, *Phytochemistry*, 27, 1145 (1988).
- 12. M.C. Chamy, M. Piovano, J.A. Garbarino, V. Gambaro, and C. Miranda, Phytochemistry, 28, 571 (1989).
- 13. M. Piovano, M.C. Chamy, J.A. Garbarino, and V. Gambaro, Phytochemistry, 28, 2844 (1989).
- 14. P. Imamura, A. Marsaioli, L. Barak, and E. Ruveda, Phytochemistry, 16, 1842 (1977).

Received 26 August 1991